Interactions between an Anionic Porphyrin and Cationic Surfactants in Water

Toshihiro Tominaga,* Satoshi Endoh, and Hiroaki Ishimaru Department of Applied Chemistry, Okayama University of Science, 1-1 Ridai-cho, Okayama 700 (Received October 15, 1990)

Interactions between decyl-, dodecyl-, and tetradecyltrimethylammonium bromide (DeTAB, DTAB, and TTAB) and an anionic porphyrin, 5,10,15,20-tetrakis(4-sulfonatophenyl)porphine (H₂TPPS⁴⁻), have been studied in water at 298.2 K. Precipitate formation constants between the surfactant cations and the H₂TPPS⁴⁻ anion have been obtained. The precipitates were resolubilized by a further addition of the surfactants at concentrations close to the critical micelle concentrations (cmc) in DeTAB, and below the cmc's in DTAB and TTAB solutions. Surface tension and conductance measurements show that the effects of H₂TPPS⁴⁻ on the cmc's are very small. The fluorescence of H₂TPPS⁴⁻ is quenched in both water and sodium dodecyl sulfate solutions by the methylviologen cation, but not in cationic surfactant solutions. The diffusion coefficients were measured in a system containing H₂TPPS⁴⁻ and TTAB; the results show the existence of large premicellar aggregates.

Because of their importance in biological and photochemical processes, porphyrins and metalloporphyrins have attracted much interest. Recently, watersoluble porphyrins have been shown to interact with 1,10-phenanthroline (phen) and pyridine derivatives,¹⁾ amino acids,²⁾ and other ionic species.³⁾ Dimerization⁴⁾ and counterion-induced dimerization⁵⁾ of cationic and anionic porphyrins have been studied. Photoinduced electron-transfer and charge-separation processes have been studied extensively in microheterogeneous solutions including micelles and vesicles.6-8) To understand these processes, an elucidation of the interactions of porphyrins with surfactants is essestial. Although there have been studies on the interactions between water-soluble porphyrins and polymers, 9,10) there have been no systematic studies on the interactions between ionic porphyrins and surfactants.

We report here on the interactions between an anionic porphyrin, 5,10,15,20-tetrakis(4-sulfonatophenyl)porphine (H₂TPPS⁴⁻), and cationic surfactants in water. In acidic solutions, the predominant species of the porphyrin is H₄TPPS²⁻;^{4b,9a} it tends to aggregate.^{4a,4b} We have studied systems where H₂TPPS⁴⁻ is the predominant species. Measurements were made on precipitate formation and resolubilization concentrations, surface tension, conductance, fluorescence quenching, and diffusion coefficient. In a previous paper, we reported interactions between a metal chelate cation, [Fe(phen)₃]²⁺, and anionic surfactants.¹¹⁾ The present results are compared with the previous ones.

Experimental

Materials and Preparation of Solutions. H₆TPPS (Tokyo Kasei, specially prepared analytical reagent), sodium dodecyl sulfate (NaDS) (BDH, 99.0%), decyltrimethylammonium bromide (DeTAB), dodecyltrimethylammonium bromide (DTAB), and tetradecyltrimethylammonium bromide (TTAB) (Tokyo Kasei, guaranteed reagents) were used as received. Water was distilled twice. A stock solution of H₂TPPS⁴⁻ was

prepared by neutralizing the H₆TPPS solution with NaOH. This solution was used to prepare buffered and nonbuffered solutions of the desired concentrations. The absence of H₄TPPS²⁻ was checked by the absorption spectrum, which showed a maximum at 413 nm, but no peak at 434 nm at the Soret bands. ¹²⁾ Metalloporphyrin solutions were prepared by adding a trace amount of Cd(NO₃)₂ to the solution of H₂TPPS⁴⁻ as a catalyst, ¹³⁾ and then adding equivalent amounts of the respective metal ions. The porphyrin and metalliporphyrin solutions were protected from light.

Measurements. All measurements were made at 298.2 K. For determining precipitate formation and the resolubilization concentrations, solutions of both H2TPPS4- and an alkyltrimethylammonium bromide were placed in a water thermostat controlled to ± 0.1 K. After temperature equilibration, solutions of H₂TPPS⁴⁻ and the surfactant were mixed and replaced in the bath. The solubilization limit was determined by observing the presence or absence of precipitates, or turbidity. The absorption and fluorescence spectra were measured by using a Hitachi 228A spectrophotometer and a Hitachi F-2000 fluorescence spectrophotometer, respectively. The surface tensions of surfactant solutions were measured by using a De Nouy-type tensiometer (Furukawa Seisakusho). The solution conductivities were measured at 5 kHz by means of a linear-bridge conductometer (Huso Model 360 B) in an oil thermostat controlled to ± 0.005 K.¹¹⁾

The diffusion coefficients were measured by the Taylor dispersion method. ¹⁴⁾ In this method a small amount of solution is injected into a solvent flowing through a capillary tube. The solute is dispersed along the tube. From the concentration profile at the end of the tube the difussion coefficient can be calculated. For tracer diffusion measurements, the solvent flowing through the capillary contained a specified amount of TTAB; the injected solution contained the same amount of TTAB and 10 μM (1 M=1 mol dm⁻³) H₂TPPS⁴⁻. Because the H₂TPPS⁴⁻ concentration was well below that of TTAB, the diffusion coefficient could be taken as the tracer diffusion coefficient. (Note that the concentration of the solute is further diluted as it flows through the capillary tube.) For interdiffusion between metalloporphyrins, the solvent flowing through the capillary contained a spec-

ified amount of TTAB and $50\,\mu\text{M}$ [Mn^{III}-(tpps)]³⁻; the injected solution contained the same amount of TTAB as the solvent and $50\,\mu\text{M}$ [M^{II}(tpps)]⁴⁻, where M stands for Cu or Ni. This was possible because [Mn^{III}(tpps)]³⁻ has a weak absorption at 413 nm where [Ni^{II}(tpps)]⁴⁻ and [Cu^{II}(tpps)]⁴⁻ have an intense absorption. The concentration profile of the metalloporphyrin was monitored by spectrophotometric detectors, either Waters-490 or UVIDEC 100-IV. Other details concerning the measurements have been described. ^{11,15)}

Results and Discussion

Precipitate Formation and Resolubilization. When a cationic surfactant was added to a fixed concentration of H₂TPPS⁴⁻, a precipitate appeared, which was then resolubilized by a further addition of the surfactant. Figure 1 shows a phase diagram for precipitate formation and resolubilization for TTAB.

Regions A and C correspond to clear solutions and region B corresponds to turbid (two-phase) systems. A similar phase diagram has been shown for the [Fe(phen)₃]²⁺-DS⁻ system. ¹⁶⁾ In an ammonium buffer (pH=8.0, I (ionic strength) = 0.1 M), precipitate formation concentrations were larger and the resolubilization concentrations lower than in water. Figure 2 shows the effect of the surfactant chain length. With increasing surfactant chain length, both precipitate formation and resolubilization concentrations decrease. In Figs. 1 and 2, the cmc's of the surfactants determined from surface tension measurements are shown. For DTAB and TTAB, the resolubilization of the precipitate occurs at surfactant concentrations below their cmc's. This will be discussed later; we first examine the precipitate formation process. precipitate formation reaction may be written as

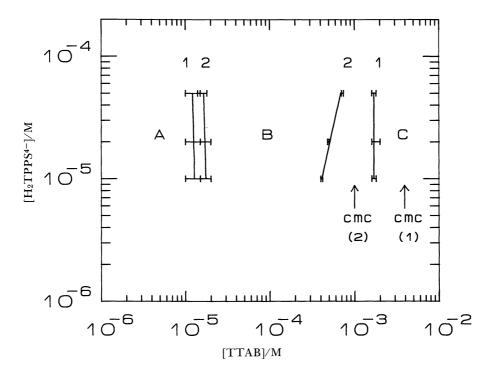


Fig. 1. Phase diagram for the precipitate formation and resolubilization for H₂TPPS⁴- and TTAB at 298.2 K. Regions A and C correspond to the one-phase clear solutions and region B corresponds to the two-phase system. 1, In water (without added salt); 2, in ammonium buffer (pH=8.0, *I*=0.1 M). Arrows indicate cmc's of TTAB in water and in the ammonium buffer.

Table 1. Values of log K for the Precipitate Formation Reaction between C_nTA⁺ and H₂TPPS⁴⁻ (Eq. 1) in Water at 298.2 K

Surfactant	$\log K$		
	<i>I</i> =0.1 M ^{a)}	No added salt	No added salt ^{b)}
$C_{10}TAB$	18.19±0.12	19.82±0.17	13.37 ± 0.36
$C_{12}TAB$	20.49 ± 0.15	21.83 ± 0.21	18.13 ± 0.19
$C_{14}TAB$	23.71 ± 0.38	24.32 ± 0.45	22.20 ± 0.34

a) Ammonium buffer (pH=8). b) The values for the reaction between C_nTA^+ and $[Fe(bpso)_3]^{4-}$ (Ref. 17).

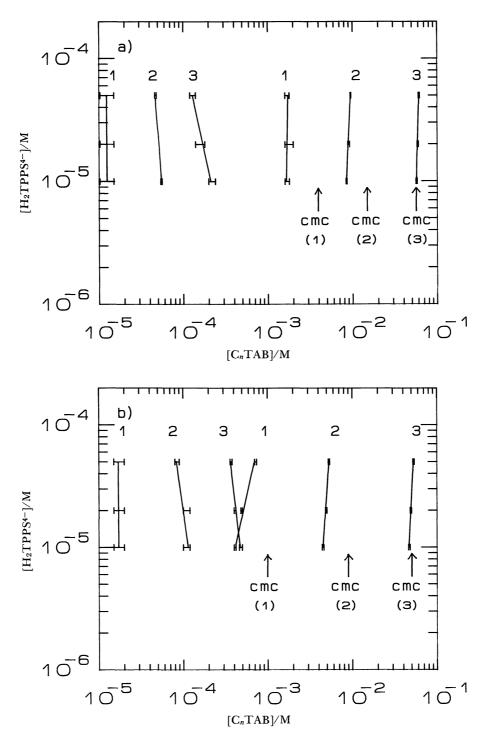


Fig. 2. Phase diagram for the precipitate formation and resolubilization for H₂TPPS⁴⁻ and TTAB (1), DTAB (2), and DeTAB (3) in water (without added salt) (a), and in ammonium buffer (pH= 8.0, *I*=0.1 M) (b) at 298.2 K. Arrows indicate cmc's of the respective surfactants.

$$4C_nTA^+ + H_2TPPS^{4-} \stackrel{K}{\rightleftharpoons} (C_nTA)_4 \cdot H_2TPPS, \qquad (1)$$

Table 1 lists the values of $\log K$. The standard Gibbs energy of transfer from the aqueous state to the solid state can be calculated as

where C_nTA^+ stands for DeTA+ (n=10), DTA+ (n=12), or TTA+ (n=14), and $K=1/\{[C_nTA^+]^4[H_2TPPS^{4-}]\}$.

$$\Delta G_{\text{ad-solid}}^{\circ} = -RT \ln K, \tag{2}$$

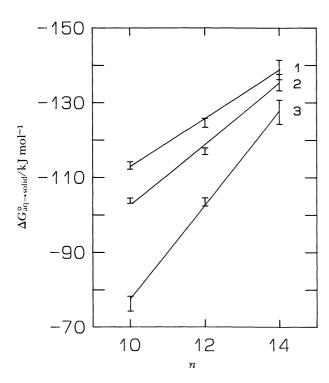


Fig. 3. Standard Gibbs energy of transfer of $(C_nTA)_4$ · H_2TPPS and $(C_nTA)_4$ · $[Fe(bpso)_3]$ from the aqueous state to the solid state at 298.2 K. 1, $(C_nTA)_4$ · H_2TPPS in water (without added salt); 2, $(C_nTA)_4$ · H_2TPPS in ammonium buffer (I=0.1 M); 3, $(C_nTA)_4$ · $[Fe(bpso)_3]$ in water (without added salt).

where R is the gas constant. In Fig. 3, the $\Delta G_{\rm aq\to solid}^{\circ}$ values are plotted against the surfactant chain length. The results may be regarded as being linear within experimental uncertainties. From the slopes, the values for $\Delta G_{\rm aq\to solid}^{\circ}$ per methylene group can be obtained, and are -6.4 and -7.9 kJ mol⁻¹ in the cases of no added salt and I=0.1 M (ammonium buffer, pH=8), respectively. Also shown in Fig. 3 is the result for [Fe(bpso)₃]⁴– (bpso=4.7-diphenyl-1,10-phenanthroline-disulfonate).¹⁷⁾ The $\Delta G_{\rm aq\to solid}^{\circ}$ value for the methylene group for this system is -12.8 kJ mol⁻¹; it is twice as large as that for the H₂TPPS⁴– case. This difference reflects differences in the standard Gibbs energy in the two kinds of solids.

Surface Tension and Conductivity. The cmc values shown in Figs. 1 and 2 were determined from the break point of the surface tension vs. concentration curve. They are 57 mM, 16 mM, and 4.0 mM in water, and 50 mM, 9.0 mM, and 1.0 mM in the ammonium buffer for DeTAB, DTAB, and TTAB, respectively. Resolubilization of the precipitate occurs at surfactant concentration below their cmc's for DTAB and TTAB, though it occurs close to the cmc's for DeTAB. In order to determine whether the cmc's decrease in the presence of H₂TPPS⁴⁻, surface tension measurements were made for DTAB and TTAB solutions in the presence of 5×10⁻⁵ M H₂TPPS⁴⁻. The results showed that

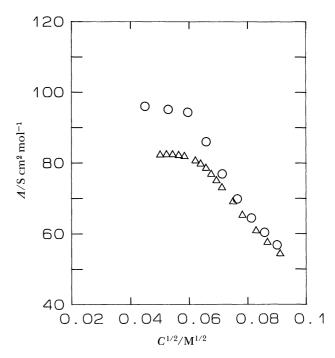


Fig. 4. Molar conductivities of TTAB in water (O) and in 0.1 mM $\rm H_2TPPS^{4-}$ solution (Δ) as a function of the TTAB concentration at 298.2 K.

the presence of H₂TPPS⁴⁻ had negligible effects on the cmc values. Figure 4 shows the molar conductivities of TTAB in the presence (0.1 mM) and absence of H₂TPPS⁴⁻. From the break point, the cmc of TTAB in water is found to be 3.8 mM, which is close to the value obtained from the surface-tension measurement (4.0 mM). As can be seen from the figure, the break point is not lowered, even in the presence of 0.1 mM H₂TPPS⁴⁻. These results suggest that, at surfactant concentrations below the cmc's and above the resolubilization concentrations, there is no "normal" micelle, but there are premicellar aggregates which may be expressed as (TTA)_x(H₂TPPS)_yBr_z(x-4y-z)+. Premicellar aggregates have been known for other ionic dyes, and are sometimes called "dye-rich induced micelles." ^{18–20}

Fluorescence Quenching. In order to obtain information about premicellar aggregates, fluorescence quenching experiments were carried out. If H₂TPPS⁴⁻ is associated with cationic surfactants, a cationic quencher may have difficulty in approaching the H₂TPPS⁴-In addition to cationic surfactant solutions, measurements were also made in water and in NaDS solutions; the results are shown in Fig. 5. In the case of no surfactant, the plot for the quenching of H₂TPPS⁴⁻ (maximum at 640 nm) by the methylviologen ion (MV²⁺) does not give a straight line. This reflects the fact that there are ion pairs between MV2+ and the ground state H₂TPPS4-, and that static quenching in addition to dynamic quenching ocurrs.6) In fact, Schmehl and Whitten have obtained a value of 647 M⁻¹ for the ion-pair formation constant between

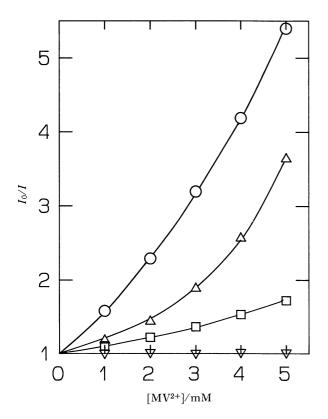


Fig. 5. Stern-Volmer plots for the quenching of H₂TPPS⁴⁻ by MV²⁺ in 0.1 M ammonium buffer (pH=8.0). O, No surfactant; Δ, in 0.02 M NaDS; □, in 0.1 M NaDS; ∇, in 0.0005 M TTAB solutions at 298.2 K.

H₂TPPS⁴⁻ and MV²⁺ in 0.05 M NaCl.⁶⁾ In NaDS (0.02 M NaDS+0.05 M NaCl), although most of the MV²⁺ ions are bound to the micelles (>98%), they have estimated that the contribution to quenching is in the following order: static quenching>dynamic quenching by free MV²⁺> dynamic quenching by micelle-bound MV^{2+,6)} This makes plots in NaDS nonlinear also (Fig. 5). In 8 mM NaDS, the results were close to those in water (not shown in Fig. 5). With increasing NaDS concentration, the degree of quenching becomes smaller (Fig. 5), since more MV²⁺ ions are bound to the micelles.

In the case of 0.5 mM TTAB, fluorescence (maximum at 647 nm) is not quenched at all. It is unlikely that there are fewer free MV²⁺ ions in the TTAB solutions than in the NaDS solutions, since MV²⁺ ions must be less bound to TTA+ ions than to NaDS micelles. The result therefore suggests that H₂TPPS⁴⁻ ions are associated with TTA+ ions, and that the overall charge of an aggregate is positive. This prevents MV²⁺ ions from comming close to the H₂TPPS⁴⁻ ions and, as a result, electron transfer from the exited H₂TPPS⁴⁻ ion to the MV²⁺ ion does not occur. The results for 2 and 20 mM TTAB (above the cmc) and 10 mM DTAB (close to the cmc) also showed no quenching (not shown in Fig. 5).

Diffusion Coefficient. The final step in this study was to obtain information about the size of the aggregates. Figure 6 shows the tracer diffusion coefficients for H₂TPPS⁴⁻ and the interdiffusion

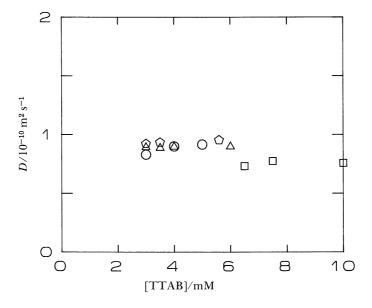


Fig. 6. Diffusion coefficients of H₂TPPS^{4−}, metalloporphyrins, and micelles in TTAB solutions at 298.2 K. △, Tracer diffusion coefficients of H₂TPPS^{4−}; △, interdiffusion coefficients of [Ni^{II}(tpps)]^{4−} and [Mn^{III}(tpps)]^{3−}; ○, interdiffusion coefficients of [Cu^{II}(tpps)]^{4−} and [Mn^{III}(tpps)]^{3−}; □, tracer diffusion coefficients of the TTAB micelles.

coefficients for pairs of metalloporphyrins in TTAB solutions. When the TTAB concentration is zero, the tracer diffusion coefficient of the H2TPPS4- ion is $2.8\times10^{-10}\,\mathrm{m^2\,s^{-1}}$ (not shown in Fig. 6), which was estimated from the limiting molar conductivity using the Nernst equation.²¹⁾ In 3—6 mM TTAB solutions, the tracer diffusion coefficient for H₂TPPS⁴⁻ is $(0.94\pm0.01)\times10^{-10}$ m² s⁻¹, and is significantly low when compared to the value in the absence of TTAB. The interdiffusion coefficient between [MII(tpps)]4-(M=Cu or Ni) and [Mn^{III}(tpps)]³⁻ in 3—6 mM TTAB is $(0.89\pm0.02)\times10^{-10}$ m² s⁻¹, and is close to the tracer diffusion coefficient for H2TPPS4-.22) These values are compared with the tracer diffusion coefficient for the TTAB micelle, $(0.75\pm0.02)\times10^{-10}$ m² s⁻¹ at 6.5—10 mM, which was obtained by measuring the diffusion of pyrene solubilized in the micelle.

By a comparison of the diffusion coefficients of the aggregate and the micelle (Fig. 6), the value of x must be large but smaller than 70, which is the agreegation number of the TTAB micelle.23) In the case of a 3 mM TTAB solution, the mole ratio of TTA+ to H₂TPPS⁴⁻ is 300 at the time of injection, and increases with time for tracer diffusion measurements, while the mole ratio of TTA+ to $[M(tpps)]^{n-}$ (n=3 or 4) is 60 throughout the measurement for the metalloporphyrin interdiffusion cases. In the tracer diffusion case, therefore, there are sufficient TTA+ ions to bind with the H₂TPPS⁴⁻ ion and form large aggregates. For the interdiffusion case, however, the maximum value of x is 60, where all TTA+ ions are bound to $[M(tpps)]^{n-1}$ ions. This suggests that the value of y is larger than one. Such kinds of premicellar aggregates are, as mentioned before, sometimes called "dye-rich induced micelles". 18-20) As we have seen, the tracer diffusion coefficients of H2TPPS4- and the interdiffusion coefficients of the metalloporphyrins are very close in magnitude. This suggests that the size of the premicellar aggregates of H₂TPPS⁴⁻, [M^{II}(tpps)]⁴⁻, and [Mn^{III}(tpps)]³⁻ are similar.

Comparison with Related Studies. Schmehl and Whitten have found that precipitate formation and resolubilization occurs for H₂TPPS⁴⁻ and hexadecyltrimethylammonium chloride (CTAC).⁶⁾ They have mentioned that, in the presence of H₂TPPS⁴⁻, micellization of CTAC occurs significantly below the cmc of CTAC in the absence of added ions. Although we have not studied CTAC, our results suggest that in the presence of H₂TPPS⁴⁻ although the cmc is not significantly changed (i.e., there is no normal micelle), the premicellar aggregates, each having at least one H₂TPPS⁴⁻ ion, exist below the cmc.

In a previous study, we measured the tracer diffusion coefficients for the [Fe(phen)₃]²⁺ ion in anionic surfactant solutions.¹¹⁾ In solutions of sodium 1-dodecanesulfonate (NaDSO), no precipitate forms between [Fe(phen)₃]²⁺ and DSO⁻, at least when the

[Fe(phen)₃]²⁺ concentration is lower than 2 mM and the DSO- concentration is lower than 7 mM. The tracer diffusion coefficient of [Fe(phen)₃]²⁺ (which is the tracer diffusion coefficient of the aggregate $Na_z[Fe(phen)_3]_x(DSO)_y(y-2x-z)-)$ decreased monotonically with increasing concentration of NaDSO. When NaDS is added to a fixed concentration of [Fe(phen)₃]²⁺, precipitates of [Fe(phen)₃](DS)₂ form, which are resolubilized upon further addition of NaDS below its cmc. In 3—8 mM NaDS (below the cmc), the tracer diffusion coefficient of [Fe(phen)₃]²⁺ (which is the tracer diffusion coefficient of the aggregate Na_z[Fe(phen)₃]_x $(DS)_{\nu}^{(\nu-2x-z)-}$) was lower than the tracer diffusion coefficient of the NaDS micelle. Although the cases of H₂TPPS⁴⁻ and TTAB are similar to that of [Fe(phen)₃]²⁺ and NaDS, in that in both systems a precipitate forms which is then resolubilized below cmc's of the surfactants, the tracer diffusion of H₂TPPS⁴⁻, i.e., that of the $(TTA)_x(H_2TPPS)_yBr_z^{(x-4y-z)+}$ aggregates, is not smaller than that of the tracer diffusion coefficient of the TTAB micelles.

It would be of interest to see if the diffusion coefficient of the premicellar aggregates between [Fe(bpso)₃]⁴⁻ and TTAB is larger or smaller than that of the TTAB micelles. However, the resolubilization of the (TTA)₄[Fe(bpso)₃] precipitates occurs at a TTAB concentration close to its cmc,¹⁷⁾ and we have not been able to tell the difference in the diffusion coefficients of the premicellar aggregates and the micelles.

The authors wish to thank Dr. Masaaki Tabata of Saga University for his helpful discussions. This work was partially supported by a Grant-in Aid for Scientific Research on Priority Area of "Molecular Approaches to Non-equilibrium Processes in Solutions" No. 02245101 from the Ministry of Education, Science and Culture.

References

- 1) J. A. Shelnutt, *Inorg. Chem.*, **22**, 2535 (1983); K. Kawamura, H. Kasahara, S. Igarashi, and T. Yotsuyanagi, *Chem. Lett.*, **1987**, 2365.
- 2) M. Tabata and M. Tanaka, *Inorg. Chem.*, **27**, 203; 3190 (1988); M. Tabata and A. Shimada, *Bull. Chem. Soc. Jpn.*, **63** 672 (1990).
- 3) T. Sato, T. Ogawa, and K. Kano, *J. Phys. Chem.*, **88**, 3678 (1984); K. Kano, T. Nakajima, and S. Hashimoto, *J. Phys. Chem.*, **91**, 6614 (1987); M. Tabata and I. Yamada, *Bull. Chem. Soc. Jpn.*, **60**, 3525 (1987).
- 4) a) A. Corsini and O. Herrmann, *Talanta*, **33**, 335 (1986); b) E. B. Fleisher, J. M. Palmer, T. S. Srivastava, and A. Chatterjee, *J. Am. Chem. Soc.*, **93**, 3162 (1971); c) M. Krishnamurthy, J. R. Sutter, and P. Hambright, *J. Chem. Soc.*, *Chem. Commun.*, **1975**, 13; d) K. Kano, M. Takei, and S. Hashimoto, *J. Phys. Chem.*, **94**, 2181 (1990).
- 5) T. K. Chandrashekar, H. van Willigen, and M. H. Ebersole, J. Phys. Chem., 88, 4326 (1984); P. Firman, R. G.

- Wilkins, and S. P. Kasprzyk, J. Am. Chem. Soc., 111, 4990 (1989).
- 6) R. H. Schmehl and D. G. Whitten, *J. Phys. Chem.*, **85**, 3473 (1981).
- 7) J. K. Hurst, L. Y. C. Lee, and M. Gratzel, *J. Am. Chem. Soc.*, **105**, 7048 (1983).
- 8) T. Nagamura, N. Takeyama, K. Tanaka, and T. Matsuo, *J. Phys. Chem.*, **90**, 2247 (1986); A. Uehata, H. Nakamura, S. Usui, and T. Matsuo, *J. Phys. Chem.*, **93**, 8197 (1989).
- 9) a)F. M. El Torki, P. J. Casano, W. F. Reed, and R. H. Schmehl, *J. Phys. Chem.*, **91**, 3686 (1987); b) M.D. Yanuck and R.H. Schmehl, *Chem. Phys. Lett.*, **122**, 133 (1985).
- 10) J. Bielec, B. Pilas, T. Sarna, and T. G. Truscott, J. Chem. Soc., Faraday Trans. 2, **82**, 1469 (1986).
- 11) T. Tominaga, T. Maruyama, T. Koshiba, and Y. Yamamoto, Bull. Chem. Soc. Jpn., 62, 1745 (1989).
- 12) When TTAB was added to an acidic solution containing H₄TPPS²⁻, the absorption peak at 434 nm disappeared and the peak at 416—417 nm (2—4 mM TTAB), which corresponds to H₂TPPS⁴⁻, appeared. It is worth noting that the total charge of the porphyrin increases in spite of the fact that the microscopic dielectric constant around the porphyrin decreases as a result of the interaction with TTAB. This means that what is important is microscopic charge of the central porphyrin ring, but not the total charge of the porphyrin. Similar result has been reported for the interaction of H₄TPPS²⁻ with poly(*N*-vinylpyrrolidone).^{9a}
- 13) M. Tabata and M. Tanaka, J. Chem. Soc., Dalton Trans., 1955 (1983); M. Tanaka, Pure Appl. Chem., 55, 151 (1983)
- 14) G. I. Taylor, *Proc. R. Soc. London, Ser. A*, **219**, 186 (1953); **225**, 473 (1954); H. J. V. Tyrrell and K. R. Harris,

- "Diffusion in Liquids. A Theoretical and Experimental Study," Butterworths, London (1984), p. 193.
- 15) T. Tominaga, S. Matsumoto, T. Koshiba, and Y. Yamamoto, J. Chem. Soc., Faraday Trans. 1, 84, 4261 (1988).
- 16) D. Lemordant and R. Gaboriaud, Fluid Phase Equilib., 20, 269 (1985).
- 17) K. Osafune, T. Maruyama, and T. Tominaga, 58th National Meeting of the Chemical Society of Japan, April 1989, Abstr., No. 3IIB30.
- 18) P. Mukerjee and K. J. Mysels, *J. Am. Chem. Soc.*, **77**, 2937 (1955).
- 19) J. H. Baxendale and M. A. J. Rodgers, *Chem. Phys. Lett.*, **72**, 424 (1980); *J. Phys. Chem.*, **86**, 4906 (1982).
- 20) Y. Kusumoto and H. Sato, Chem. Phys. Lett., 68, 13 (1979); H. Sato, M. Kawasaki, and K. Kasatani, J. Phys. Chem., 87, 3759 (1983); H. Sato, M. Kawasaki, M. Haga, K. Kasatani, T. Ban, H. Suenaga, and N. Kitamura, Nippon Kagaku Kaishi, 1984, 51.
- 21) R. A. Robinson and R. H. Stokes, "Elctrolyte Solutions," 2nd ed, Butterworth, London (1970), Chap. 11.
- 22) If the Charges of the interdiffusing pair of metalloporphyrins are the same, the interdiffusion coefficient may be regared as the intradiffusion coefficient under the assumption that the diffusion coefficient is insensitive to the central metal ion.^{11,15)} However, we were unable to find a pair of tetraanionic metalloporphyrins which have sufficiently different absorption spectra so that the interdiffusion coefficient can be measured. For the definition of intradiffusion coefficient, see; R. Mills, *J. Phys. Chem.*, **69**, 3116 (1965).
- 23) P. Lianos and R. Zana, J. Colloid Interface Sci., **84**, 100 (1981); D. D. Miller, L. J. Magid, and D. F. Evans, J. Phys. Chem., **94**, 5921 (1990).